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Types and prevalence of genetic disorders in humans	<ul style="list-style-type: none"> <li>• Single gene disorders <ul style="list-style-type: none"> <li>○ There are many, but most are very rare (&lt;1 in 10,000)</li> <li>○ Total incidence in newborns is approx. 1 in 100</li> <li>○ Specific disorders may occur more frequently in certain populations</li> <li>○ Maintained due to heterozygote advantage or founder effect</li> <li>○ Most show recessive inheritance (one working copy is as good as two)</li> <li>○ Some are dominant due to: <ul style="list-style-type: none"> <li>▪ Haploinsufficiency – loss of function in a dosage sensitive gene resulting in 50% of normal protein production not being enough to produce the wild type phenotype</li> <li>▪ Dominant-negative mutations – loss of function mutations in which the protein is made but not functional, and inhibits the function of the normal protein in heterozygotes</li> <li>▪ Gain of function mutations – new function of the gene product, or the protein is always active, or there are increased levels of expression, or inappropriate expression</li> </ul> </li> </ul> </li> <li>• Chromosomal <ul style="list-style-type: none"> <li>○ Most caused by aneuploidy</li> <li>○ One chromosome is present more or less than normal</li> <li>○ More severe for larger, autosomal chromosomes</li> <li>○ Aneuploids very frequent among spontaneous miscarriages (40-50% in the first trimester)</li> <li>○ Chromosomal aberrations also occur – translocations, deletions, duplications</li> </ul> </li> <li>• Multifactorial <ul style="list-style-type: none"> <li>○ Part genetic, part environmental origin</li> <li>○ Genetic component is usually polygenic (&lt;1 gene)</li> <li>○ Congenital abnormalities (e.g. spina bifida) or late onset (e.g. type I diabetes)</li> </ul> </li> </ul>
Gene structure and expression	<ul style="list-style-type: none"> <li>• A gene is a sequence of DNA that is required for the production of a functional product – either a polypeptide or a functional RNA molecule</li> <li>• Include the coding sequence as well as adjacent sequences required for proper expression (e.g. promoters, terminators, regulatory sequences)</li> </ul>
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Determining inheritance patterns	<ul style="list-style-type: none"> <li>• Unaffected families who may carry the disease but have no affected children won't come to genetic centres, thus won't be included in data when analysed – thus may not show in a Mendelian ratio</li> <li>• Variable expressivity – may be difficult to assess the phenotype at the extreme closest to normal, especially for behavioural phenotypes (e.g. autism)</li> <li>• Variable penetrance – some individuals with the affected genotype do not show the phenotype</li> <li>• New mutations may arise</li> <li>• Locus heterogeneity – mutations in several different genes may show the same phenotype (often when mutations are in genes in biochemical pathways)</li> </ul>

Linkage analysis	<ul style="list-style-type: none"><li>Linkage groups correspond with individual chromosomes</li><li>Genes may be mapped by following their joint segregation patterns in pedigrees</li><li>Can also be mapped by association with individual chromosomes in somatic cell hybrids between humans and another species (human chromosomes variably lost)</li><li>Can also be mapped by in situ hybridisation of a labelled antisense copy, binds to its locus in a chromosome spread</li><li>Requires allelic differences at two loci</li><li>Linkage phase<ul style="list-style-type: none"><li>Coupling: <math>\frac{A\ B}{a\ b}</math></li><li>Repulsion: <math>\frac{A\ b}{a\ B}</math></li></ul></li><li>The chance of crossover occurring between two loci is proportional to their distances apart</li><li>Maximum value is 50% (above this and the markers are on different chromosomes)</li></ul>																												
LOD scores	<ul style="list-style-type: none"><li>Allows sufficient data to be collected from pedigrees (won't be able to get enough offspring to analyse from one family when using human data)</li><li>Based on calculating the chance of getting a sibship assuming the two loci show a recombination fraction of <math>r</math> (range 0-0.5)<ul style="list-style-type: none"><li>Divide this by the chance of getting the sibship assuming the two loci are unlinked (<math>R=0.5</math>)</li><li>This is the odds that the genes are linked (<math>\theta</math>)</li><li>Take <math>\log_{10}</math> of this (<math>Z</math>), called the LOD score (log of the odds)</li><li>Repeat for all sibships</li><li>Then add the individual LOD scores (because they are logs, they can be added)</li></ul></li><li>E.g. if 7 offspring all show parental inheritance</li></ul> <div><p><b>Pedigree 1:</b></p><table><tr><th><math>r</math></th><th>chance <math>(1 - r)^7</math></th><th>odds (<math>\theta</math>) chance <math>r</math>/(chance <math>r = 0.5</math>)</th><th><math>\log_{10}</math> odds (<math>Z</math>) LOD</th></tr><tr><td>0.0</td><td>1.000</td><td>128</td><td>2.11</td></tr><tr><td>0.1</td><td>0.478</td><td>61.2</td><td>1.79</td></tr><tr><td>0.2</td><td>0.210</td><td>26.8</td><td>1.43</td></tr><tr><td>0.3</td><td>0.082</td><td>10.5</td><td>1.02</td></tr><tr><td>0.4</td><td>0.028</td><td>3.58</td><td>0.554</td></tr><tr><td>0.5</td><td>0.0078</td><td>1.00</td><td>0.000</td></tr></table></div> <ul style="list-style-type: none"><li>Highest odds indicate the most likely</li><li>Z score must be higher than 3</li></ul>	$r$	chance $(1 - r)^7$	odds ( $\theta$ ) chance $r$ /(chance $r = 0.5$ )	$\log_{10}$ odds ( $Z$ ) LOD	0.0	1.000	128	2.11	0.1	0.478	61.2	1.79	0.2	0.210	26.8	1.43	0.3	0.082	10.5	1.02	0.4	0.028	3.58	0.554	0.5	0.0078	1.00	0.000
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Identifying human disease genes	<ul style="list-style-type: none"><li>Positional cloning<ul style="list-style-type: none"><li>Identifying a gene after mapping it</li><li>The more extensive the mapping, the smaller the candidate region, the easier it is</li></ul></li><li>Steps in positional cloning<ul style="list-style-type: none"><li>Obtain the sequence of all the DNA in the candidate region</li><li>Identify all the genes in the region</li><li>Prioritise the genes for mutant screening (obtain candidate genes)</li><li>Test candidate genes for mutations in affected individuals</li></ul></li></ul>																												

	<ul style="list-style-type: none"> <li>○ Confirm through animal models</li> <li>● Obtaining the DNA sequence – chromosome walking <ul style="list-style-type: none"> <li>○ Genomic DNA subjected to partial restriction digest</li> <li>○ Generates large fragments of overlapping DNA sequences</li> <li>○ Cloned into a vector – makes a genomic library</li> <li>○ Start by identifying a clone that overlaps one of the markers that identify the candidate region</li> <li>○ Use this first clone to probe the library and identify overlapping DNA fragments</li> </ul> </li> <li>● Identifying all the genes in the region <ul style="list-style-type: none"> <li>○ Gene prediction – look for open reading frames</li> <li>○ Zoo blots – probe southern blot of genomic DNA from other species with human probes (sequences part of genes are more likely to be conserved)</li> <li>○ CpG islands – clusters of unmethylated CpG dinucleotides found near many transcription initiation sites</li> <li>○ Exon trapping – clone random fragments from the region of interest into a special vector that is engineered so that a splicing reaction will occur if the cloned fragment contains an intron/exon boundary</li> </ul> </li> <li>● Prioritise the genes to obtain candidate genes <ul style="list-style-type: none"> <li>○ Perform BLAST searches with predicted genes</li> <li>○ Look for appropriate expression</li> </ul> </li> <li>● Confirming the candidate gene <ul style="list-style-type: none"> <li>○ Mutation screening in affected individuals</li> <li>○ Rescue the phenotype</li> <li>○ Production of an animal model of the disease</li> </ul> </li> </ul>
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Human mutation	<ul style="list-style-type: none"> <li>● Types of single gene mutations <ul style="list-style-type: none"> <li>○ Base substitution – single base change <ul style="list-style-type: none"> <li>▪ Transition – pyrimidine replaced by another pyrimidine (C/T) or purine by purine (G/A)</li> <li>▪ Transversion – purine replaced by pyrimidine (or vice versa)</li> </ul> </li> <li>○ Insertions or deletions – short DNA sequences may be deleted or added</li> </ul> </li> <li>● Effects of single gene mutations on gene products <ul style="list-style-type: none"> <li>○ Within coding sequence <ul style="list-style-type: none"> <li>▪ Silent mutations – no alteration to amino acid sequence (production may be slowed down)</li> <li>▪ Missense mutations – amino acid does change</li> <li>▪ Nonsense mutation – codon changes to a stop codon</li> <li>▪ Frameshift mutation – insertion or deletion of 1-2 nucleotides, changing the reading frame</li> </ul> </li> <li>○ In non-coding regions <ul style="list-style-type: none"> <li>▪ Promoter regions – may increase or decrease transcription</li> <li>▪ Splice recognition sites – pre-mRNA may not be spliced correctly</li> <li>▪ 5'UTR/3'UTR – alteration in ability of mRNA to be translated or in mRNA stability</li> </ul> </li> </ul> </li> <li>● Effects of a mutation on gene function <ul style="list-style-type: none"> <li>○ Null/amorph – produces no product</li> <li>○ Hypomorph – produces a reduced amount or activity or</li> </ul> </li> </ul>